

Hands-On Session: Regression Analysis

File: [afni05_regression.pdf](#)

SSCC/NIMH/NIH/HHS



Overview

- What we have learned so far
 - Use data viewer 'afni' interactively
 - Model HRF with a **fixed-shape** basis function (e.g., Gamma variate)
 - Assume the brain responds with the **same shape**
 - Across **subjects** and any activated **regions**
 - Differ in **magnitude**: β (and its significance) is focus
- What we will do in this hands-on session
 - Data pre-processing overview for time series regression analysis
 - Basic concepts
 - Regressors, design matrix, and confounding effects
 - Statistical significance testing in regression analysis
 - Navigation with GUI '**afni**'
 - Spot check for the original data
 - Statistic thresholding with data viewer '**afni**' (**two-sided** vs. **one-sided** with t)
 - Model performance (visual check of curve fitting and test via full F or R^2)

FMRI Regression Analysis

- Voxel-wise regression model: $y = X\beta + \varepsilon$
 - y : signal (time series) at a voxel - **different** across voxels
 - X : explanatory (independent) variables (regressors) - **same** across voxels
 - β : regression coefficients (response strength) - **different** across voxels
 - ε : residuals (anything we can't account for) - **different** across voxels
- Regressors in design matrix $X = [x_1, x_2, \dots, x_k]$
 - Regressors of interest: hemodynamic responses (HDR)
 - Regressors of no interest: drift effect (polynomials), head motion, *etc.*
- Association between stimulus and BOLD signal: HDR/HRF
 - Fixed shape regardless of subjects, brain regions, stimuli: regression
 - No assumption about the HDR shape: deconvolution + regression
 - Middle ground: regression
- Residuals
 - White noise: OLS - 3dDeconvolve
 - Serially correlated: ARMA(1,1)+REML - 3dREMLfit

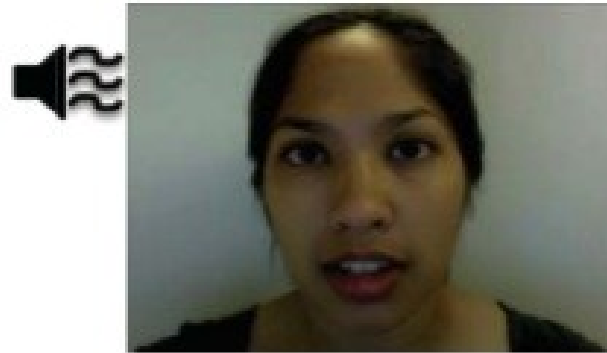
A Case Study

- ◆ **Speech Perception Task:** Subjects were presented with audiovisual speech presented in a predominantly auditory or predominantly visual modality.
- ◆ A digital video system was used to capture auditory and visual speech from a female speaker.
- ◆ 2 types of stimulus conditions:



(1) **Auditory-Reliable**

Example: Subjects can clearly *hear* the word “cat,” but the video of a woman mouthing the word is degraded.

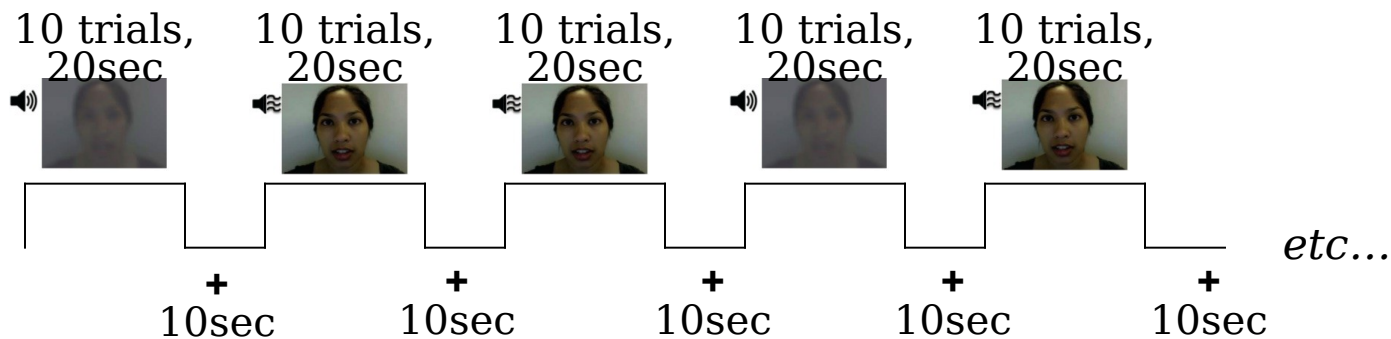


(2) **Visual-Reliable**

Example: Subjects can clearly *see* the video of a woman mouthing the word “cat,” but the audio of the word is degraded.

Experiment Design

- ◆ 3 runs in a scanning session
- ◆ Each run consisted of randomized 10 blocks:
 - 5 blocks contained Auditory-Reliable (*Arel*) stimuli, and
 - 5 blocks contained Visual-Reliable (*Vrel*) stimuli
- ◆ Each block contained 10 trials of *Arel* OR *Vrel* stimuli
 - Each block lasted for 20s (1s for stimulus presentation, followed by a 1s inter-stimulus interval)
- ◆ Each baseline block consisted of a 10s fixation point

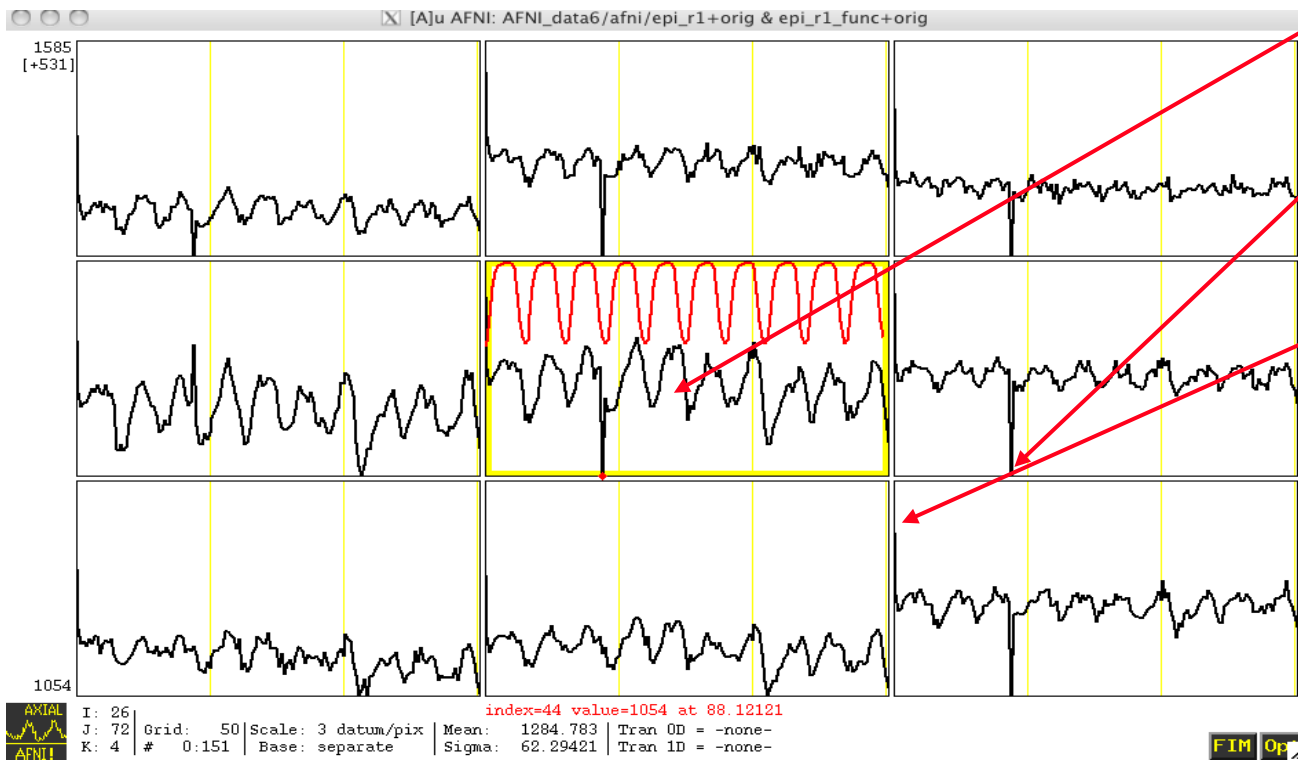


Data Collected

- ◆ 2 anatomical datasets for each subject from a 3T
 - 175 sagittal slices
 - voxel dimensions = $1.0 \times 0.938 \times 0.938 \text{ mm}^3$
- ◆ 3 time series (EPI) datasets for each subject
 - 33 axial slices \times 152 volumes (TRs) per run
 - TR = 2s; voxel dimensions = $2.75 \times 2.75 \times 3.0 \text{ mm}^3$
- ◆ Sample size, $\underline{n} = 10$ (all right-handed subjects)
 - 10 is far too few for most modern experiments

Data Quality Check

- To look at the data: type **cd AFNI_data6/afni**, then **afni**
- **Switch Underlay** to dataset **epi_r1**
 - Then **Axial Image** and **Graph**
 - **FIM→Pick Ideal** ; then click **afni/epi_r1_ideal.1D** ; then **Set**
 - Right-click in image, **Jump to (ijk)**, then **26 72 4**, then **Set**



- Data clearly has activity in sync with reference
 - 20s blocks
- Data also has a big spike at 89s
 - Head motion
- Spike at $t = 0$
- Some tricks with keyboard
 - **a**: automatic scaling
 - **v**: video mode
 - **m/M**: voxel matrix sizing on Graph window

Preparing Data for Analysis

- Following preparatory steps are common (e.g., `afni_proc.py`):
 - Outliers: **`3dToutcount`**, **`3dDespike`**
 - Temporal alignment or slice timing correction (sequential/interleaved): **`3dTshift`**
 - EPI Image/volume registration (head motion correction): **`3dvolreg`**
 - EPI to anatomy registration: **`align_epi_anat.py`**
 - Spatial normalization (standard space conversion): **`@auto_tlrc`**, **`auto_warp.py`**
 - Blurring/smoothing: **`3dmerge`**, **`3dBlurToFWHM`**, **`3dBlurInMask`**
 - Masking: **`3dAutomask`**
 - Temporal mean scaling: **`3dTstat`** and **`3dcalc`**
- Not all steps are necessary or desirable in any given case

Regression Analysis

- Regression model: $y = X\beta + \varepsilon$
- Run script by typing **tcsh rall_regress** (takes a few minutes)

```

3dDeconvolve -input rall_vr+orig -polort 1 \
  -concat '1D: 0 150 300' \
  -num_stimts 8 \
  -stim_times 1 stim_AV1_vis.txt 'BLOCK(20,1)' -stim_label 1 Vrel \
  -stim_times 2 stim_AV2_aud.txt 'BLOCK(20,1)' -stim_label 2 Arel \
  -stim_file 3 motion.1D'[0]' -stim_base 3 -stim_label 3 roll \
  -stim_file 4 motion.1D'[1]' -stim_base 4 -stim_label 4 pitch \
  -stim_file 5 motion.1D'[2]' -stim_base 5 -stim_label 5 yaw \
  -stim_file 6 motion.1D'[3]' -stim_base 6 -stim_label 6 dS \
  -stim_file 7 motion.1D'[4]' -stim_base 7 -stim_label 7 dL \
  -stim_file 8 motion.1D'[5]' -stim_base 8 -stim_label 8 dP \
  -gltsym 'SYM: Vrel -Arel' -glt_label 1 V-A \
  -tout -x1D rall_X.xmat.1D -xjpeg rall_X.jpg \
  -fitts rall_fitts -bucket rall_func \
  -jobs 2

```

- 2 audiovisual stimulus classes were given using **-stim_times**
- **Important to include motion parameters as regressors?**
 - › May remove the confounding effects due to motion artifacts
 - › 6 motion parameters as covariates via **-stim_file + -stim_base**
 - › **motion.1D** generated from **3dvolreg** with the **-1Dfile** option
 - › Test the significance of head motion parameters
 - › Add **-bout** or remove **-stim_base**
 - › Use **-gltsym 'SYM: roll \ pitch \ yaw \ dS \ dL \ dP'**

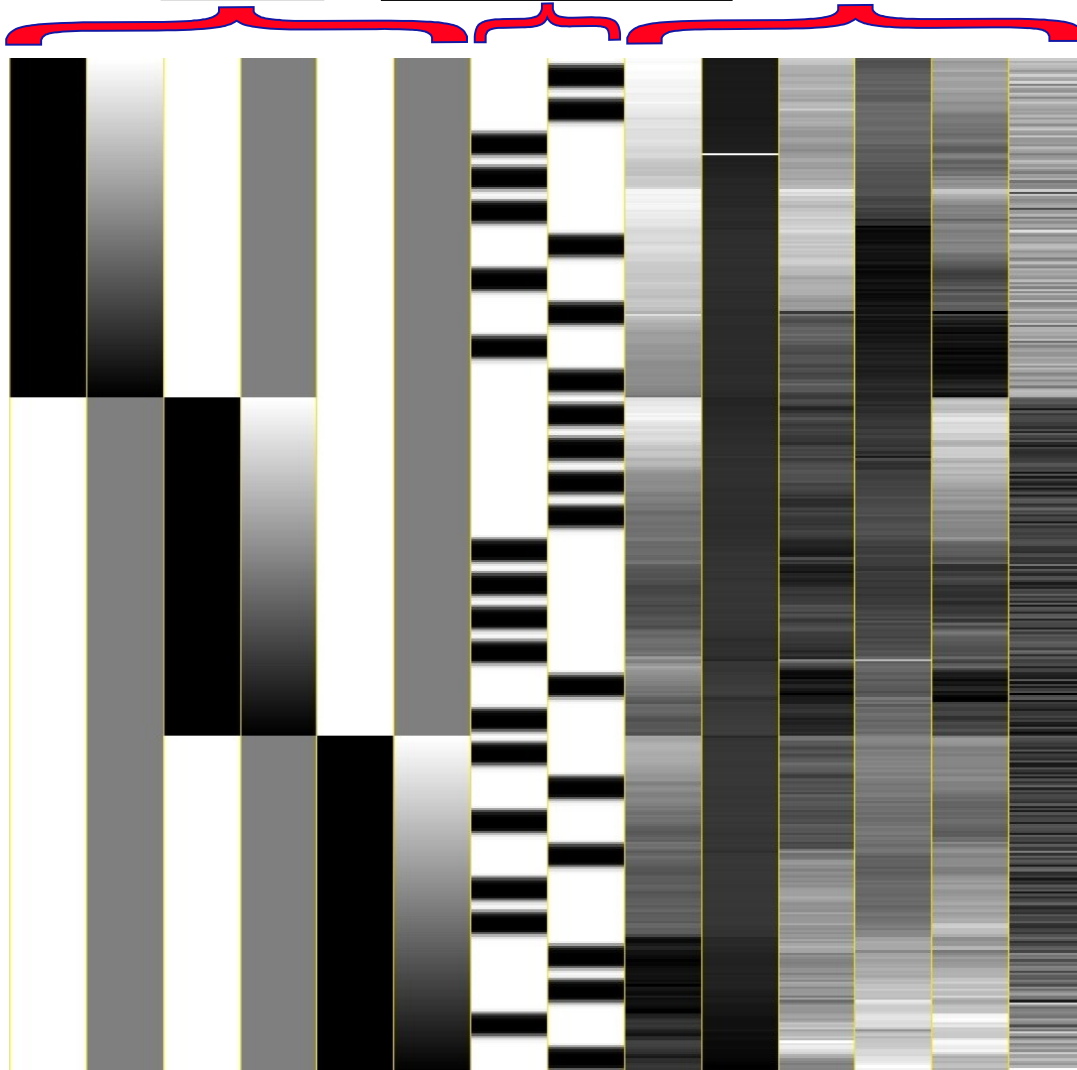
Modeling Serial Correlation in the Residuals

- Temporal correlation exists in the residuals of the time series regression model
- Within-subject variability (or statistical value) would get deflated (or inflated) if temporal correlation is not accounted for in the model
- Better correct for the temporal correlation if bringing both effect size and within-subject variability to group analysis
- ARMA(1, 1) assumed in **3dREMLfit**
- Script automatically generated by 3dDeconvolve (may use `-x1D_stop`)
 - ★ File **rall_func.REML_cmd** under **AFNI_data6/afni**
 - ★ Run it by typing ***tcs*h -x *rall_func.REML_cmd***

```
3dREMLfit -matrix rall_X.xmat.1D -input rall_vr+orig \  
-tout -Rbuck rall_func_REML -Rvar rall_func_REMLvar \  
-Rfitts rall_fitts_REML -verb
```

Regressor Matrix X for This Script (via -xjpeg)

Baseline Audiovisual stimuli Head Motion

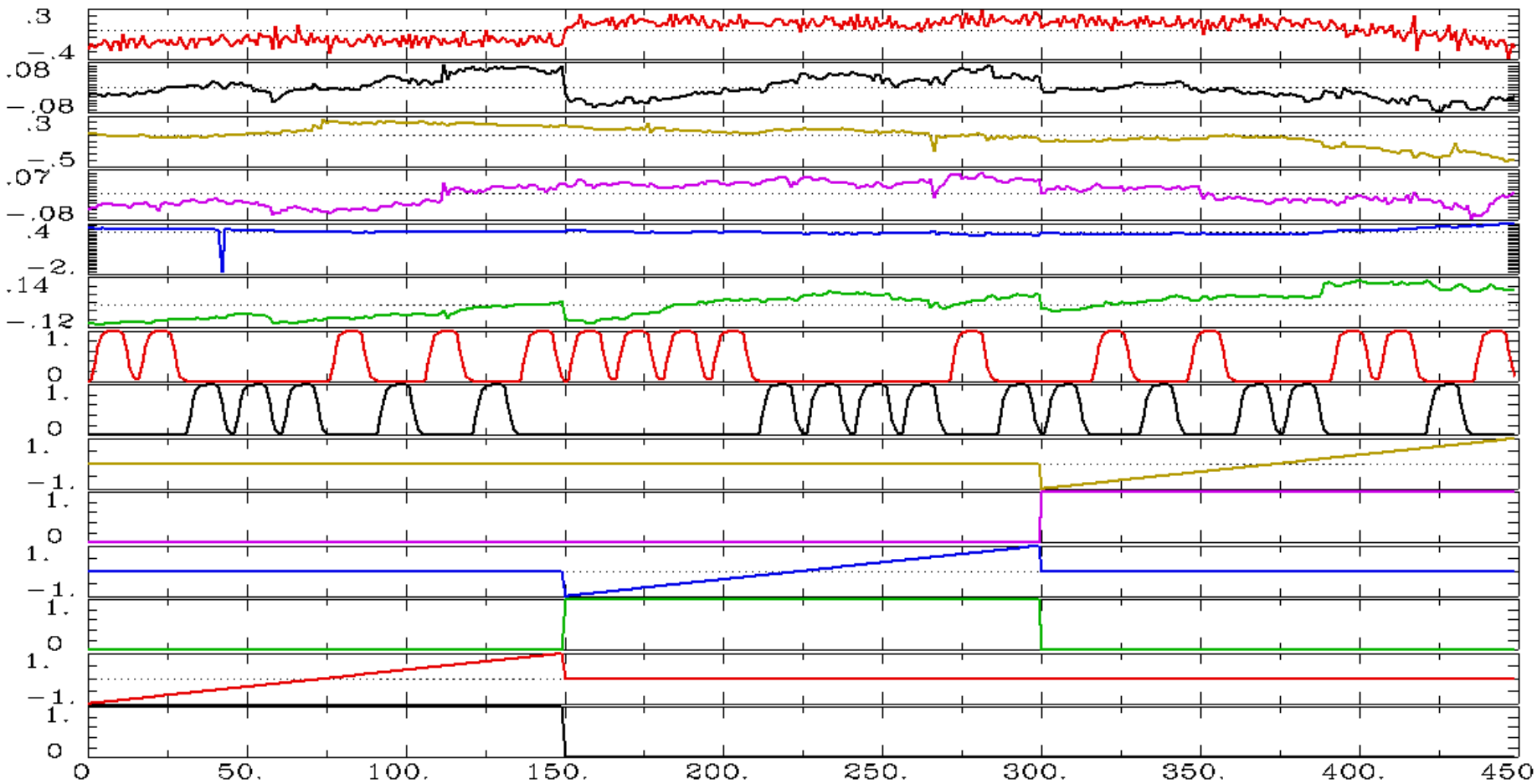


- 6 drift effect regressors
 - linear baseline
 - 3 runs times 2 params/run
- 2 regressors of interest
- 6 head motion regressors
 - 3 rotations and 3 shifts

`aiv_rall_xmat.jpg`

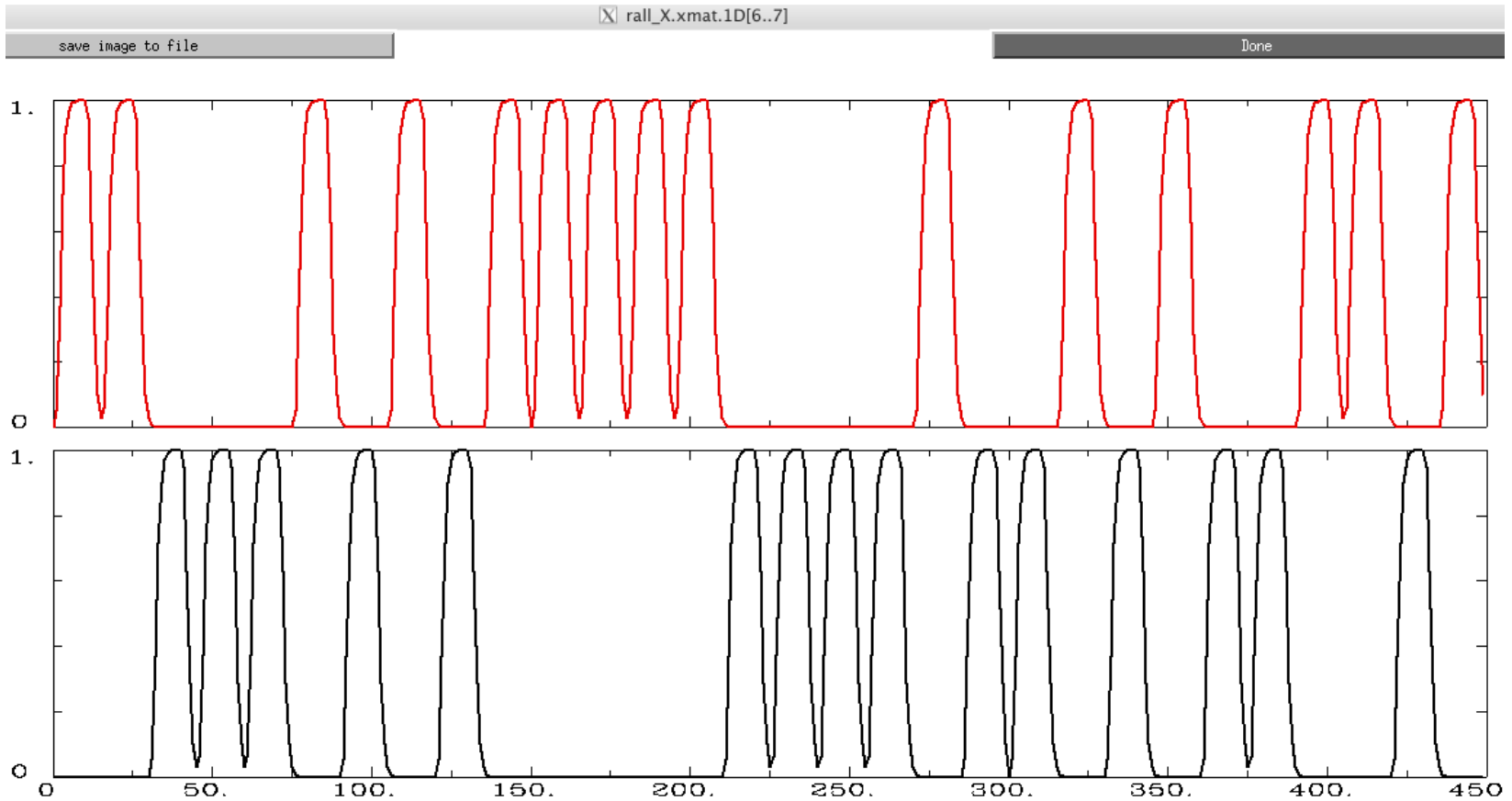
Showing All Regressors (via -x1D)

save image to file Done



All regressors: **1dplot -sepscl rall_X.mat.1D**

Plotting Regressors of Interest



Regressors of Interest: `1dplot rall_X.mat.1D'[6..7]'`

Options in 3dDeconvolve - 1

-concat '1D: 0 150 300'

- “File” that indicates where distinct imaging runs start inside the input file
 - Numbers are the time (TR) **indexes** inside the dataset file for start of runs
 - These time points are considered as **discontinuities** in the model
 - In this case, a text format .1D file put directly on the command line
 - Could also be a filename, if you want to store that data externally

-num_stimts 8

- 2 audiovisual stimuli (+6 motion), thus 2 **-stim_times** below
- Times given in the **-stim_times** files are *local* to the start of each run

-stim_times 1 stim_AV1_vis.txt 'BLOCK(20,1)' -stim_label 1 Vrel

• Content of **stim_AV1_vis.txt**

```
60 90 120 180 240
120 150 180 210 270
0 60 120 150 240
```

★ Each of 3 lines specifies start time in **seconds** for stimuli within the run

Options in 3dDeconvolve - 2

```
-gltsym 'SYM: Vrel -Arel' -glt_label 1 V-A
```

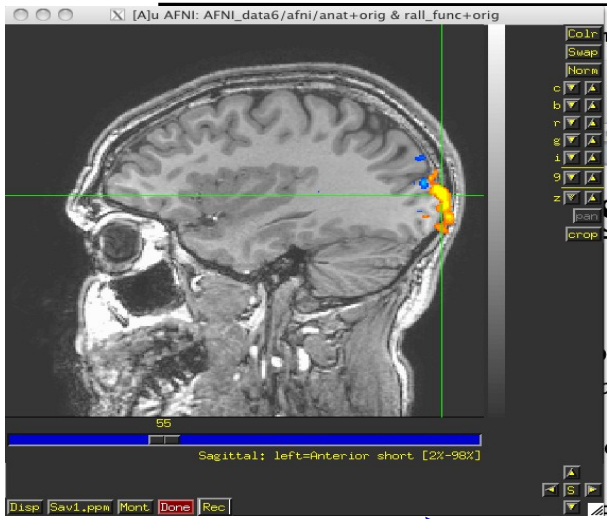
- **GLTs**: **G**eneral **L**inear **T**ests
- **3dDeconvolve** provides test statistics for each regressor separately, but to test combinations of the β weights in each voxel, we need **-gltsym** option
- Example above tests the difference between the β weights for the **Virtual-reliable** and the **Audio-reliable** responses
 - **SYM**: means symbolic input is on command line
 - Otherwise inputs will be read from a file
 - Symbolic names for each regressor taken from **-stim_label** options
 - Stimulus label can be preceded by **+** or **-** to indicate sign to use in combination of β weights
 - **Leave space after each label!**
- Goal is to test a linear combination of the β weights
 - Null hypothesis $\beta_{Vrel} = \beta_{Arel}$
 - e.g., does **Vrel** get different response from **Arel**?
- What do **'SYM: 0.5*Vrel +0.5*Arel'** and **'SYM: Vrel \ Arel'** test?

Options in 3dDeconvolve - 3

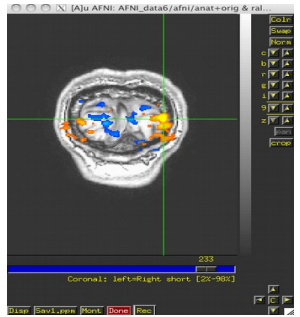
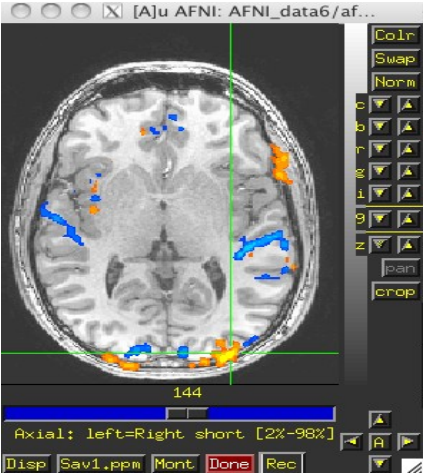
-fout -tout = output both F - and t -statistics for each stimulus class (**-fout**) and stimulus coefficient (**-tout**) — but not for the baseline coefficients (use **-bout** for baseline)

- The full model statistic is an F -statistic that shows how well all the regressors of interest explain the variability in the voxel time series data
 - Compared to how well *just* the baseline model time series fit the data times (in this example, we have 12 baseline regressor columns in the matrix — 6 for the linear drift, plus 6 for motion regressors)
 - $F = [SSE(r) - SSE(f)] / df(n) \div [SSE(f) / df(d)]$
- The individual stimulus classes also will get individual F - (if **-fout** added) and/or t -statistics indicating the significance of their individual *incremental* contributions to the data time series fit
 - If $DF=1$ (e.g., F for a single regressor), t is equivalent to F : $t(n) = F^2(1, n)$

Results of **rall_regress** Script



The AFNI control panel for the 'rall_regress' script. It includes a coordinate display (UI=3633129, 01=69973), view selection (Axial, Sagittal, Coronal), and a 'Define Overlay' button. A T-t intensity scale is visible on the right, ranging from -1.00 to 1.00, with a current value of 2.757. The 'Clusters' section shows 'Ulay #0 #0' and 'Olay #6 V-A_GLT#0_Tstat'.



The AFNI menu for selecting an overlay. The list includes: #0 Full_Fstat, #1 Vrel#0_Coef, #2 Vrel#0_Tstat, #3 Arel#0_Coef, #4 Arel#0_Tstat, #5 V-A_GLT#0_Coef, and #6 V-A_GLT#0_Tstat. The 'Set' button is highlighted in red.

• Images showing results from third GLT contrast: **VrelvsArel**

• Menu showing labels from **3dDeconvolve**

• Play with these results yourself!

Compare 3dDeconvolve and 3dREMLfit

Terminal — afni — 74x30

```

You have mail.
gangc@fingol:~> cd AFNI_data6/afni
gangc@fingol:~/AFNI_data6/afni> afni &
[1] 40244
gangc@fingol:~/AFNI_data6/afni>
Thanks go to LR Frank for useful feedback

GPL AFNI: Analysis of Functional NeuroImages, by RW Cox (rwcox@nih.gov)
This is Version AFNI_2009_12_31_1431
[[Precompiled binary macosx_10.6_Intel_64: Mar 18 2010]]

** This software was designed to be used only for research purposes. **
** Clinical uses are not recommended, and have never been evaluated. **
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**** If you DO find AFNI useful, please cite this paper:
RW Cox. AFNI: Software for analysis and visualization of
functional magnetic resonance neuroimages.
Computers and Biomedical Research, 29:162-173, 1996.

**** If you find SUMA useful, citing this paper also would be nice:
ZS Saad, RC Reynolds, B Argall, S Japee, RW Cox.
SUMA: An Interface For Surface-Based Intra- And Inter-Subject Analysis
With AFNI. 2nd IEEE International Symposium on Biomedical Imaging:
Macro to Nano 2, 1510-1513, 2004.

Initializing: X11.

```

Group Analysis: will be carried out on β or GLT coef (+t-value) from single-subject analysis